



Bilirubin Encephalopathy

Shuo Qian¹ · Prateek Kumar¹ · Fernando D. Testai¹

Accepted: 12 April 2022 / Published online: 19 May 2022

© The Author(s), under exclusive licence to Springer Science+Business Media, LLC, part of Springer Nature 2022

Abstract

Purpose of Review Hyperbilirubinemia is commonly seen in neonates. Though hyperbilirubinemia is typically asymptomatic, severe elevation of bilirubin levels can lead to acute bilirubin encephalopathy and progress to kernicterus spectrum disorder, a chronic condition characterized by hearing loss, extrapyramidal dysfunction, ophthalmoplegia, and enamel hypoplasia. Epidemiological data show that the implementation of universal pre-discharge bilirubin screening programs has reduced the rates of hyperbilirubinemia-associated complications. However, acute bilirubin encephalopathy and kernicterus spectrum disorder are still particularly common in low- and middle-income countries.

Recent Findings The understanding of the genetic and biochemical processes that increase the susceptibility of defined anatomical areas of the central nervous system to the deleterious effects of bilirubin may facilitate the development of effective treatments for acute bilirubin encephalopathy and kernicterus spectrum disorder. Scoring systems are available for the diagnosis and severity grading of these conditions. The treatment of hyperbilirubinemia in newborns relies on the use of phototherapy and exchange transfusion. However, novel therapeutic options including deep brain stimulation, brain-computer interface, and stem cell transplantation may alleviate the heavy disease burden associated with kernicterus spectrum disorder.

Summary Despite improved screening and treatment options, the prevalence of acute bilirubin encephalopathy and kernicterus spectrum disorder remains elevated in low- and middle-income countries. The continued presence and associated long-term disability of these conditions warrant further research to improve their prevention and management.

Keywords Hyperbilirubinemia · Bilirubin encephalopathy · Bilirubin-induced neurotoxicity · Kernicterus spectrum disorder

Introduction

Bilirubin is an endogenous yellowish pigment that results from the metabolism of the heme group contained in red blood cells (Fig. 1). Neonatal hyperbilirubinemia is common and mostly benign, manifesting with jaundice which is typically observed in the first week of life. Based on observational studies, it has been estimated that 60% of term and 80% of preterm neonates experience hyperbilirubinemia [1, 2]. The term bilirubin encephalopathy (BE) refers to the acute and chronic neurological dysfunction seen in association with

severe hyperbilirubinemia. Extremely elevated levels of total bilirubin ($TB \geq 25$ mg/dL) can lead to acute bilirubin encephalopathy (ABE) and cause significant morbidity or mortality if not adequately treated. Survivors often experience long-term neurologic deficits ranging from mild movement disorders and/or auditory dysfunction to severe choreoathetoid cerebral palsy, auditory neuropathy, and gaze palsies [3]. In the next sections, we discuss the epidemiology, clinical features, pathogenesis, and treatments of BE.

Terminology and Epidemiology

Several terms have been adopted to describe bilirubin neurotoxicity including BE, ABE, kernicterus, chronic bilirubin encephalopathy, and bilirubin-induced neurological dysfunction (BIND). The term “kernicterus” has been used interchangeably for both acute and chronic bilirubin-induced neurotoxicity since its conception in 1903 by pathologist

This article is part of the Topical Collection on *Neurology of Systemic Diseases*

✉ Shuo Qian
sqian7@uic.edu

¹ Department of Neurology and Rehabilitation, University of Illinois at Chicago College of Medicine, 912 S Wood St, Chicago, IL 60612, USA

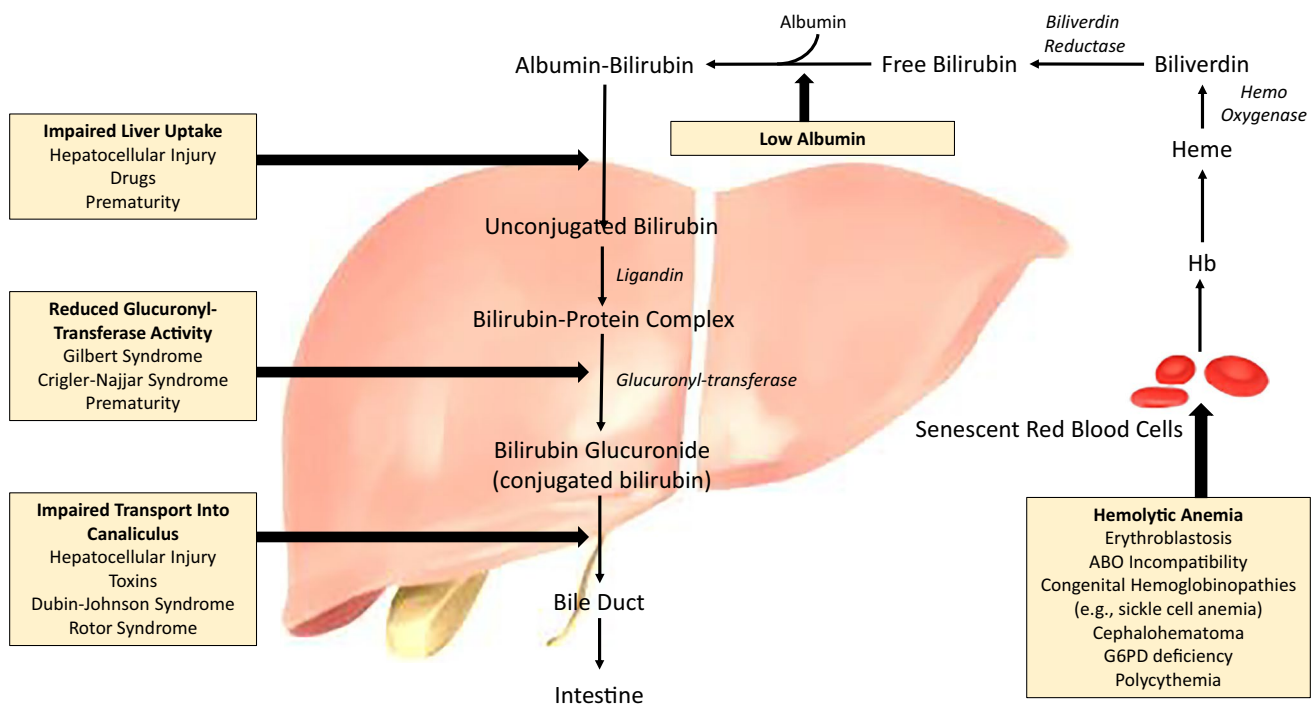


Fig. 1 Metabolism of bilirubin. Bilirubin originates primarily from the senescence of circulating red blood cells. Heme, a protoporphyrin ring that functions as a prosthetic group in hemoglobin (Hb), is sequentially metabolized to biliverdin and bilirubin. In physiologic conditions, more than 99% of the unconjugated bilirubin in the circulation is bound to albumin and only a small portion remains as

free bilirubin which is liposoluble and can cross the blood–brain barrier. In the liver, bilirubin is conjugated with glucuronic acid which is water soluble and excreted into the biliary track and the intestine. Hyperbilirubinemia in the neonate can result from overproduction of bilirubin (hemolytic anemia), low availability of albumin, reduced UDP-glucuronyltransferase activity, or impaired excretion

Christian Schmorl, who used it to describe the post-mortem findings of yellow staining of the basal ganglia (BG), hippocampus, and central parts of the cerebellum [4]. To avoid confusion, in 2004, the American Academy of Pediatrics (AAP) recommended to use the term ABE for the manifestations of bilirubin neurotoxicity seen in the first weeks after birth, and “kernicterus” for the chronic and permanent clinical sequelae [5]. Le Pichon JB et al. further suggested to replace all the terms previously used for chronic BE with “kernicterus spectrum disorder” (KSD) to encompass the gamut of long-term neurological sequelae resulting from BE [6]. In this review, we use ABE and KSD to describe the acute- and long-term consequences of bilirubin neurotoxicity, respectively, and BE as a more generic term that encompasses ABE and KSD.

The epidemiology of BE varies significantly across the globe and the population investigated. In general, the prevalence for kernicterus in high-income countries is estimated as 10/100,000 live births. However, the implementation of universal pre-discharge bilirubin screening programs and the early treatment of hyperbilirubinemia has reduced the rate of complications associated with it. As an example, data from the Healthcare Cost and

Utilization Project Kids’ Inpatient Database (KID) for the years 1997–2012 show that the proportion of infants diagnosed with hyperbilirubinemia increased from 9.4 to 15.5% during the study period. In parallel, the rate of discharges with the diagnosis of kernicterus decreased from 7 to 1.9 per 100,000 newborns [7]. In comparison, the rate of kernicterus in low- and middle-income countries (LMIC) has remained particularly elevated, with a prevalence of approximately 73/100,000 live births reported in Eastern Europe/Central Asia, Latin America, sub-Saharan Africa, and South Asia [8].

Risk factors for neurotoxicity from extreme hyperbilirubinemia include central nervous system (CNS) prematurity, hemolytic disease due to blood group incompatibility, inherited red blood cell enzymatic deficiencies or membrane defects (e.g., glucose-6-phosphate dehydrogenase deficiency), genetic predisposition, and medical conditions such as sepsis, intracranial hemorrhage, and acidosis (Fig. 1) [9, 10]. There are also multiple reports describing delayed presentation of BE without a history of ABE during the first days of life which suggests that additional unknown risk factors may play a role in the disease.

Clinical Manifestations

ABE has been categorized into three phases with variable onset. Phase 1 or early ABE is typically seen during the first 1–3 days of the illness and is characterized by nonspecific symptoms of decreased alertness, poor feeding and sucking, mild hypotonia, and hyperreflexia. Phase 2 or intermediate ABE usually presents by the end of the first week with stupor, irritability, and hypertonia of extensor muscles resulting in opisthotonos, retrocollis, and high-pitched cry. Phase 3, also known as late or advanced ABE, is seen after the first week and is characterized by hypotonia or hypertonia, pronounced opisthotonos-retrocollis and high-pitched crying, apnea, deep stupor or coma, fever, and sometimes seizures [11•, 12•].

KSD, which refers to the chronic and enduring clinical consequences of bilirubin toxicity, may develop slowly over years in some children that survive ABE. During the first year, decreased muscle tone, increased deep tendon reflex, delayed achievement of developmental milestones and motor skills, and persistent tonic neck reflex are present [12•]. After the first year, the clinical manifestations are often variable and classically represented by the tetrad of hearing loss, motor (extrapyramidal) dysfunction, ophthalmoplegia, and enamel hypoplasia. Auditory and motor symptoms are the most salient features leading to disability. Damage of auditory networks manifests as auditory neuropathy spectrum disorder (ANSD) [13]. This is one of the earliest features of KSD and can often be the only clinical manifestation. Rather than sensorineural dysfunction, ANSD is characterized by absent or diminished auditory brainstem responses, sometimes accompanied by auditory nerve dysfunction sparing the cochlea and hair cells [14–17]. Children suffering from ANSD may have pure tone thresholds ranging from mild to profound hearing loss and experience reduced speech perception, leading to an inability to perform sound localization or speech discrimination [18, 19]. Extrapyramidal dysfunction presents largely as dystonia and choreoathetosis [20]. The presentation of movement disorders is variable but the upper extremities are typically more severely affected. Rapid and jerky movements, tremors, ballismus, and dystonic posturing are common [12•]. Visual abnormalities include upward gaze paresis, horizontal gaze dysfunction, and a blank stare or “scared” appearance resulting from the upward gaze paresis and facial dystonia. Dental abnormalities are relatively uncommon and include dental enamel hypoplasia and green-stained teeth [21]. Cognitive function is typically preserved in children with KSD.

Pathophysiology

BE results from the deposition of unconjugated bilirubin (UCB), particularly free bilirubin (B_f) in specific anatomical areas of the CNS, including the brainstem nuclei, BG, thalamus, hippocampus, and cerebellum [22, 23, 24•]. The

mechanism of bilirubin-induced neurotoxicity is incompletely understood. However, post-mortem and animal studies suggest the active role of neuroinflammation, excitotoxicity, mitochondrial failure, and DNA damage [11•, 25].

Hyperbilirubinemia itself results from the imbalance of the production and excretion of bilirubin. Infants have high turnover of red blood cells [26] with limited ability to conjugate bilirubin. While most neonatal hyperbilirubinemias are physiologic and rarely exceed 5 to 6 mg/dL, pathologic jaundice may occur due to increased production of bilirubin, deficiency of hepatic uptake, impaired conjugation of bilirubin, or increased enterohepatic circulation of bilirubin [21]. This can lead to the accumulation of UCB and B_f , the latter being a lipid-soluble compound that can cross the blood–brain barrier (BBB) [15]. B_f has a cytotoxic effect on astrocytes and neurons, resulting in microglial and astrocyte activation, impaired myelination, and neuronal death [27–29]. Its accumulation in the CNS and cerebrospinal fluid is mediated by transporters in the choroid plexus epithelia, endothelial cells, astrocytes, and neurons [30]. B_f increases significantly when UCB approaches the maximum binding capacity of albumin (1 g of albumin can bind 8 mg of bilirubin, ~ 1:1 on a molar basis) [31]. Whether bilirubin-induced neurotoxicity occurs or not depends on different variables, including serum UCB and albumin levels, bilirubin binding capacity by albumin, concentration of hydrogen ions, BBB function, neuronal susceptibility, and degree of CNS maturity, as well as the duration and severity of the hyperbilirubinemia [32–34].

Proposed molecular mechanisms of bilirubin-induced neurotoxicity involve excessive glutamate release, upregulation of proinflammatory cytokines, mitochondrial failure, and DNA damage [11•, 25]. Bilirubin-induced impairment of membrane transporters in astrocytes and neurons causes an increase and prolonged presence of the excitatory neurotransmitter glutamate in the synaptic cleft [35]. Cytokines such as tumor necrosis factor alpha and interleukin-1 β released by astrocytes may be involved in this process [36]. Excessive glutamate results in overstimulation of neuronal N-methyl D-aspartate (NMDA) receptors causing an increased influx of sodium, calcium, chloride, and water, generating free radicals and triggering cell death by necrosis or apoptosis [37–40]. UCB can also interact with nerve cell membranes causing oxidative damage and increased permeability, compromising lipoprotein structure and eventually disrupting cell homeostasis [41]. Furthermore, UCB can impair oxidative phosphorylation in mitochondria, leading to decreased energy production and increased mitochondrial permeability resulting in swelling and release of cytochrome C into the cytosol [30]. The subsequent activation of caspase-3 (an executioner caspase) and translocation of Bax [42] activate

mitochondrial-mediated pro-apoptotic pathways. In addition, caspase 8 (an initiator of apoptosis) may be activated as well, implicating the participation of cell-surface death receptor pathways [43]. Novel mechanisms have been more recently described. To this end, using a mouse model of neonatal hyperbilirubinemia, Rawat V et al. described that DNA injury via bilirubin-induced oxidative stress can also contribute to neuronal death and BE. In addition, it was observed that bilirubin activates main DNA repair pathways through homologous recombination and nonhomologous end joining, suggesting that these may constitute endogenous protective mechanisms against BE [25].

Data obtained in human and animal models have demonstrated that the brainstem nuclei, BG, hippocampus, and cerebellum are particularly sensitive to the detrimental effects of bilirubin [44–52]. Affected brainstem nuclei include auditory (cochlear, superior olivary, trapezoid body, lateral lemniscus, inferior colliculi), oculomotor, and vestibular nuclei [24•]. Auditory networks appear to be particularly sensitive to bilirubin toxicity and primarily involve brainstem nuclei and cranial nerve VIII. Bilirubin deposition in the BG, particularly the globus pallidus and subthalamic nuclei, as well as in the cerebellar Purkinje cells, likely causes the motor symptoms of BE [22, 48, 49]. The hippocampal CA2 region is another commonly affected structure, and the inhibition there of long-term potentiation of synaptic transmission consequently affects learning and memory [53].

Gazzin et al. conducted a study using the *jj*-sulfa Gunn rat model of hyperbilirubinemia and observed greater accumulation of UCB in the cerebellum and inferior colliculi, along with lesser accumulation in other “resistant” areas including the cortex and superior colliculi, of which functionality (cognition and vision, respectively) is usually preserved in kernicterus [54]. Several theories have been proposed to address this regional gradient in the accumulation of bilirubin. One hypothesis suggests a role of parvalbumin (PV), a calcium-binding protein [24•], due to decreased PV immunoreactivity in brainstem nuclei of dystonic *jj*-sulfa Gunn rats with severe kernicterus [50, 51]. As 95% of the neurons in the globus pallidus are GABAergic and 50–60% co-express PV, Shapiro et al. suggest that degeneration of PV-GABAergic neurons may be the most important cause of kernicterus-associated dystonia [55–58]. Another theory involves multi-drug resistance P-glycoproteins and multi-drug resistance-associated proteins (MRPs/Mrps), two large families of ATP-binding cassette transporters that mediate the export of compounds that may be toxic for the brain, including perhaps UCB [59]. There is evidence supporting the role of MRP1/Mrp1 in maintaining intracellular homeostasis and protection of CNS cells against UCB accumulation and toxicity. It has been hypothesized that variations in the expression and/or function of MRP1 may explain regional selectivity to UCB toxicity and differences in the severity of BE among neonates [30].

New technological advances have allowed the identification of genetic variations that contribute to bilirubin-induced neurotoxicity. The genes *UGT1A1* and *SLCO1B1* have been implicated in abnormal bilirubin processing [60, 61]. Also, cytochrome P450 monooxygenases (Cyps) are proteins that oxidize bilirubin. Studies have confirmed upregulated mRNA expression of *Cyp1a1*, *Cyp1a2*, and *Cyp2a3* in neurotoxicity-resistant regions of the brain as compared to more sensitive regions, suggesting a potential protective effect of these isoforms [54]. Data obtained in experimental models, in addition, show that UCD activates microglial cells which exert an early neuroprotective effect by clearing damaged neurons. In a later stage, however, microglial cells secrete pro-inflammatory mediators which have a direct effect on astrocytes. Together, microglial cells and activated astrocytes contribute to the production of additional inflammatory mediators which are cytotoxic and enhance astrogliosis. Furthermore, UCB can compromise myelinogenesis and axonal transmission directly or via microglial cells [36, 62–66]. Understanding the genetic and biochemical mechanisms that govern these responses can help us explain the different clinical manifestations observed in patients with similar bilirubin levels, and may also help us identify infants at high risk of developing permanent brain damage [24•, 67].

Diagnosis

Historically, the diagnosis of BE relied on identification of the aforementioned autopsy findings. In contemporary practice, however, the diagnosis of ABE or KSD can be made with reasonable certainty using the combination of medical history, physical examination, laboratory testing, clinical scoring systems, neurophysiological studies, and neuroimaging. Relevant history includes symptom onset and duration and history of risk factors such as CNS immaturity, hemolytic disease, family history of anemia or jaundice, and neurotoxic factors such as sepsis, acidosis, and intracerebral hemorrhage. Hyperbilirubinemia severity and duration, along with a detailed clinical examination, are also important for the diagnosis.

Jaundice alone is a suboptimal screening tool to detect hyperbilirubinemia. Thus, many countries have implemented universal screening of bilirubin levels [68]. Total plasma or serum bilirubin levels were classically determined in the laboratory by measuring the absorbance at 460 nm. Newer approaches, such as the BR2 Bilirubin Stat Analyzer from Advanced Instruments (Norwood, MA), can determine total and direct bilirubin levels in neonates using samples of only 30 μ L. In addition, noninvasive transcutaneous bilirubin (TcB) readers are now widely available in most high-income countries. Studies have shown a strong correlation between

TcB and TB measurements in both term [69, 70] and pre-term newborns [71, 72], although serum albumin levels and phototherapy may impact the accuracy of this technique [73, 74]. In addition, new point-of-care instruments such as two-color icterometers (Bilistrip™) [75] and smartphone-based methods (BiliCam) [76] can help mothers and healthcare providers make the initial determination of whether a child needs to be seen by a physician. Free bilirubin is classically measured by the “peroxidase method,” a methodology that is not widely available and considered exceedingly challenging for most clinical laboratories to perform. However, new technological advances, including the use of fluorescent sensors, may soon facilitate the accurate determination of B_f [77, 78].

Auditory involvement in BE can be assessed non-invasively by auditory brainstem responses (ABR). It has been proposed that ABR has a sensitivity of 100% and a specificity of 99.4% for detecting auditory involvement [12•]. Patients typically have abnormal or absent ABRs, with normal or giant cochlear microphonic responses and varying otoacoustic emissions [6•]. It is possible that in the near future, new technologies become available that are specifically designed to improve the diagnosis of BE and predict the risk and severity of KSD. These could include the determination of genetic risk by using a “bilirubin risk chip” or the assessment of CNS function by using fully automated neurophysiologic approaches, such as the modern “next gen” ABR technology.

Clinical scoring systems have been designed to aid in the diagnosis and grading of BE. The BIND [79] and modified BIND (BIND-M) scores (Table 1) [80] have been validated and widely adopted in clinical practice. The BIND score evaluates mental status, muscle tone, and cry characteristics to delineate three levels of ABE: subtle (1–3), moderate (4–6), and advanced (7–9). The BIND-M score additionally incorporates possible upward gaze paresis. A BIND-M score ≥ 3 was highly predictive of a clinical diagnosis of ABE, with a sensitivity of 90.7% and specificity of 97.7% [80]. Scores 1–4, 5–6, and 7–12 represent mild, moderate, and severe ABE, respectively. The BIND-M helps differentiate ABE from other causes of jaundice with similar clinical presentations. The KSD scoring system, based on the KSD Diagnostic Toolkit (KSD-TK) (Tables 2 and 3), allows a better characterization and grading of KSD [6•]. This scoring system focuses on motor and auditory disabilities as these are considered clinically significant and quantifiable features of BE. The identification and grading of clinical features such as oculomotor dysfunction can be challenging, and dental enamel dysplasia is rarely present. In a retrospective study including 37 patients, KSD-TK had 96.6% sensitivity and 87.5% specificity [6•, 81].

Finally, magnetic resonance imaging (MRI) has been shown to improve accuracy of diagnosis in BE. In ABE, MRI commonly shows bilateral T1 hyperintensity of the globus pallidus [82–84], and one study demonstrated significant correlation between bilirubin levels and apparent diffusion

Table 1 Modified bilirubin-induced neurologic dysfunction (BIND-M) score

	Score
Mental status	
• Normal	0
• Sleepy but arousable or decreased feeding	1
• Lethargy, poor suck, and/or irritable/jittery with short-term strong suck	2
• Stupor, apnea, seizures, or coma	3
Muscle tone	
• Normal	0
• Persistent mild hypotonia	1
• Moderate hypotonia, moderate hypertonia, or increasing arching of neck and trunk on stimulation without spasms of arms and legs and without trismus	2
• Persistent retrocollis, opisthotonus, or crossing or scissoring of arms or legs but without spasms of arms and legs and without trismus	3
Cry pattern	
• Normal	0
• High pitched	1
• Shrill	2
• Inconsolable crying or either weak or absent in child with previous history of high pitched or shrill cry	3
Oculomotor or eye movements	
• Normal	0
• Sun-setting or paralysis of upward gaze	3

Table 2 Kernicterus spectrum disorder (KSD) toolkit scoring system

Symptoms	Severity	Characteristics	Score
Auditory	None	No auditory symptoms	0
	Mild	ABR abnormal but present or CAPD \pm mild hearing loss; normal or mildly delayed speech	1
	Moderate	Absent or persistent abnormal ABR, mild/moderate hearing loss, may fluctuate; speech delayed or absent	2
	Severe	Absent ABR, severe-to-profound hearing loss/deafness	3
Motor	None	No motor symptoms	0
	Mild	Mild abnormal muscle tone \pm athetosis; mild gross motor delay	1
	Moderate	Moderate abnormal muscle tone \pm athetoid cerebral palsy; able to ambulate with or without assistance	2
	Severe	Severe abnormal tone \pm athetoid cerebral palsy; unable to ambulate, feed self, sign, speak; often with episodes of severe increased tone and muscle cramps	3

ANSD, auditory neuropathy spectrum disorder; ABR, auditory brainstem response; CAPD, central auditory processing disorder

Table 3 Kernicterus spectrum disorder (KSD) severity and type

		Auditory symptoms or severity			
		None	Mild	Moderate	Severe
Motor symptoms or severity	None	None	Mild auditory	Moderate auditory	Severe auditory
	Mild	Mild motor	Mild motor and auditory	Mild motor, moderate auditory	Mild motor, severe auditory
	Moderate	Moderate motor	Moderate motor, mild auditory	Moderate motor and auditory	Moderate motor, severe auditory
	Severe	Severe motor	Severe motor, mild auditory	Severe motor, moderate auditory	Severe motor and auditory

coefficients (ADC) [85]. In 2021, Wu et al. showed that the accuracy of the diagnosis of ABE can be improved by using deep learning algorithms involving multimodal MRI with the combination of T1- and T2-weighted imaging, ADC, and convolutional neural networks [86]. In comparison, children with KSD often show increased T2 signals of the globus pallidus, especially in cases having significant motor symptoms [15, 87, 88].

Management

Phototherapy and exchange transfusion (ET) are the mainstay of treatment for hyperbilirubinemia in newborns. Phototherapy, which has been used for over half a century to treat newborns with jaundice, reduces the levels of UCB regardless of its etiology [89]. When exposed to light, a fraction of parenchymal and circulating bilirubin undergoes configurational isomerization (reversible), structural isomerization (irreversible), and photooxidation [90]. Though phototherapy does not address the underlying cause of hyperbilirubinemia, it can effectively lower UCB levels and ameliorate the severity of the symptoms [91]. ET is typically used for severe neonatal hyperbilirubinemia unresponsive to phototherapy [92]. Guidelines for the use of these two treatments

in term [5, 68], preterm, and low-birth-weight infants have been published elsewhere [93, 94]. In addition, the use of intravenous immune globulin has been shown to reduce the need for exchange transfusions in Rh and ABO hemolytic disease and is recommended if TB levels continue to rise despite intensive phototherapy or remain within 2 to 3 mg/dL of the exchange transfusion level [95]. Other treatment strategies, such as pharmacologic treatment using tin-mesoporphyrin [96–99], minocycline [100–102], or caffeine [103], and therapeutic hypothermia [104], may reduce the severity of BE and are currently under investigation.

Regarding treatment of long-term sequelae, patients with ANSD may benefit from strategies such signal-to-noise ratio maximization with “FM-listening” devices and sound amplification through conventional hearing aids or cochlear implants [105]. For patients with moderate to severe motor dysfunction, symptomatic treatment relies on the use of benzodiazepines, baclofen, trihexyphenidyl (anticholinergic), and tetrabenazine (antidopaminergic). Botulinum toxin injection and intrathecal baclofen are sometimes offered to patients with severe and refractory symptoms. There is anecdotal evidence supporting the use of valbenazine or cannabinoil oil [106•], but high-quality clinical trials in this area have not been conducted. Similarly, the use of deep brain stimulation targeting the globus pallidus interna [107] has

shown promising results in patients with secondary dystonia due to KSD. However, larger studies are needed to confirm the efficacy and sustainability of this approach. Other therapeutics under investigation for the treatment of severe motor dysfunction include brain-computer interface implantation [108–110] and the use of autologous stem cells [111–114].

Conclusion

More than a century has passed since the first BE case was reported. Despite ABE and KSD being preventable and treatable, the incidence of these conditions remains elevated, particularly in low- and middle-income countries. Early diagnosis and treatment of hyperbilirubinemia are the best strategies to prevent the occurrence of bilirubin-induced neurotoxicity, which can lead to long-term disability and death. Our armamentarium for the treatment of chronic neurologic complications is expanding. However, the promising results observed in small cohorts require confirmation using properly powered studies.

Compliance with Ethical Standard

Conflict of Interest Shuo Qian, Prateek Kumar, and Fernando D Testai each declare no potential conflicts of interest.

Human and Animal Rights and Informed Consent This article does not contain any studies with human or animal subjects performed by any of the authors.

References

Papers of particular interest, published recently, have been highlighted as: • Of importance

1. Ansong-Assoku B, Shah SD, Adnan M, Ankola PA. Neonatal jaundice. Treasure Island: StatPearls; 2022.
2. Mitra S, Rennie J. Neonatal jaundice: aetiology, diagnosis and treatment. *Br J Hosp Med (Lond)*. 2017;78(12):699–704. <https://doi.org/10.12968/hmed.2017.78.12.699>.
3. Shapiro SM. Chronic bilirubin encephalopathy: diagnosis and outcome. *Semin Fetal Neonatal Med*. 2010;15(3):157–63. <https://doi.org/10.1016/j.siny.2009.12.004>.
4. Orth J. Ueber das Vorkommen von Bilirubinkristallen bei neugeborenen Kindern. In: Virchows Arch Path Anat. 1875(63):S. 447–62.
5. American Academy of Pediatrics Subcommittee on H. Management of hyperbilirubinemia in the newborn infant 35 or more weeks of gestation. *Pediatrics*. 2004;114(1):297–316. <https://doi.org/10.1542/peds.114.1.297>.
6. • Le Pichon JB, Riordan SM, Watchko J, Shapiro SM. The neurological sequelae of neonatal hyperbilirubinemia: definitions, diagnosis and treatment of the kernicterus spectrum disorders (KSDs). *Curr Pediatr Rev*. 2017;13(3):199–209. <https://doi.org/10.2174/1573396313666170815100214>. **This paper clarifies terminology, proposes the term KSD, and outlines available treatment options. The authors identify diagnostic criteria and systemic nomenclature for KSD using the KSD-Toolkit clinical scoring system.**
7. Vidavalur R, Devapatla S. Trends in hospitalizations of newborns with hyperbilirubinemia and kernicterus in United States: an epidemiological study. *J Matern Fetal Neonatal Med*. 2021;1–6. <https://doi.org/10.1080/14767058.2021.1960970>.
8. Greco C, Arnolda G, Boo NY, Iskander IF, Okolo AA, Rohsiswatmo R, et al. Neonatal jaundice in low- and middle-income countries: lessons and future directions from the 2015 Don Ostrow Trieste Yellow Retreat. *Neonatology*. 2016;110(3):172–80. <https://doi.org/10.1159/000445708>.
9. Dong XY, Wei QF, Li ZK, Gu J, Meng DH, Guo JZ, et al. Causes of severe neonatal hyperbilirubinemia: a multicenter study of three regions in China. *World J Pediatr*. 2021;17(3):290–7. <https://doi.org/10.1007/s12519-021-00422-3>.
10. • Olusanya BO, Osibanjo FB, Slusher TM. Risk factors for severe neonatal hyperbilirubinemia in low and middle-income countries: a systematic review and meta-analysis. *PLoS One*. 2015;10(2):e0117229. <https://doi.org/10.1371/journal.pone.0117229>. **This systematic review and meta-analysis examines studies from 1990 to 2014 to identify key risk factors for BE in LMIC.**
11. • Usman F DU, Shapiro SM, Le Pichon JB, Slusher TM. Acute bilirubin encephalopathy and its progression to kernicterus: current perspectives. *Res Rep Neonatol*. 2018;8:33–44. **This review describes terminology, clinical manifestations, pathogenesis, diagnosis, and treatment options for BE.**
12. • Karimzadeh P, Fallahi M, Kazemian M, Taslimi Taleghani N, Nouripour S, Radfar M. Bilirubin induced encephalopathy. *Iran J Child Neurol*. 2020;14(1):7–19. **This paper is a recent summary on the incidence, clinical manifestations, BIND scoring system, and few management options for BE.**
13. Olds C, Oghalai JS. Audiologic impairment associated with bilirubin-induced neurologic damage. *Semin Fetal Neonatal Med*. 2015;20(1):42–6. <https://doi.org/10.1016/j.siny.2014.12.006>.
14. Oghalai JS. The cochlear amplifier: augmentation of the traveling wave within the inner ear. *Curr Opin Otolaryngol Head Neck Surg*. 2004;12(5):431–8. <https://doi.org/10.1097/01.moo.0000134449.05454.82>.
15. Shapiro SM, Bhutani VK, Johnson L. Hyperbilirubinemia and kernicterus. *Clin Perinatol*. 2006;33(2):387–410. <https://doi.org/10.1016/j.clp.2006.03.010>.
16. Xia A, Song Y, Wang R, Gao SS, Clifton W, Raphael P, et al. Prestin regulation and function in residual outer hair cells after noise-induced hearing loss. *PLoS ONE*. 2013;8(12):e82602. <https://doi.org/10.1371/journal.pone.0082602>.
17. Choi CH, Oghalai JS. Perilymph osmolality modulates cochlear function. *Laryngoscope*. 2008;118(9):1621–9. <https://doi.org/10.1097/MLG.0b013e3181788d72>.
18. Shapiro SM, Nakamura H. Bilirubin and the auditory system. *J Perinatol*. 2001;21 Suppl 1:S52–5; discussion S9–62. <https://doi.org/10.1038/sj.jp.7210635>.
19. Starr A, Picton TW, Sininger Y, Hood LJ, Berlin CI. Auditory neuropathy. *Brain*. 1996;119(Pt 3):741–53. <https://doi.org/10.1093/brain/119.3.741>.
20. Farouk ZL, Muhammed A, Gambo S, Mukhtar-Yola M, Umar Abdullahi S, Slusher TM. Follow-up of children with kernicterus in Kano, Nigeria. *J Trop Pediatr*. 2018;64(3):176–82. <https://doi.org/10.1093/tropej/fmx041>.
21. Dennery PA, Seidman DS, Stevenson DK. Neonatal hyperbilirubinemia. *N Engl J Med*. 2001;344(8):581–90. <https://doi.org/10.1056/NEJM200102223440807>.

22. Hamza A. Kernicterus. *Autops Case Rep.* 2019;9(1):e2018057. <https://doi.org/10.4322/acr.2018.057>.
23. Kaplan M, Hammerman C. Understanding severe hyperbilirubinemia and preventing kernicterus: adjuncts in the interpretation of neonatal serum bilirubin. *Clin Chim Acta.* 2005;356(1–2):9–21. <https://doi.org/10.1016/j.cccn.2005.01.008>.
24. ● Riordan SM, Shapiro SM. Review of bilirubin neurotoxicity I: molecular biology and neuropathology of disease. *Pediatr Res.* 2020;87(2):327–31. <https://doi.org/10.1038/s41390-019-0608-0>. **This review summarizes current and recent advances in the understanding of the neuropathology and molecular biology of bilirubin neurotoxicity.**
25. Rawat V, Bortolussi G, Gazzin S, Tiribelli C, Muro AF. Bilirubin-induced oxidative stress leads to DNA damage in the cerebellum of hyperbilirubinemic neonatal mice and activates DNA double-strand break repair pathways in human cells. *Oxid Med Cell Longev.* 2018;2018:1801243. <https://doi.org/10.1155/2018/1801243>.
26. Brouillard RP. Measurement of red blood cell life-span. *JAMA.* 1974;230(9):1304–5.
27. Brites D, Fernandes A. Bilirubin-induced neural impairment: a special focus on myelination, age-related windows of susceptibility and associated co-morbidities. *Semin Fetal Neonatal Med.* 2015;20(1):14–9. <https://doi.org/10.1016/j.siny.2014.12.002>.
28. Brites D. Bilirubin injury to neurons and glial cells: new players, novel targets, and newer insights. *Semin Perinatol.* 2011;35(3):114–20. <https://doi.org/10.1053/j.semperi.2011.02.004>.
29. Silva RF, Rodrigues CM, Brites D. Rat cultured neuronal and glial cells respond differently to toxicity of unconjugated bilirubin. *Pediatr Res.* 2002;51(4):535–41. <https://doi.org/10.1203/00006450-200204000-00022>.
30. Ostrow JD, Pascolo L, Brites D, Tiribelli C. Molecular basis of bilirubin-induced neurotoxicity. *Trends Mol Med.* 2004;10(2):65–70. <https://doi.org/10.1016/j.molmed.2003.12.003>.
31. Cayabyab R, Ramanathan R. High unbound bilirubin for age: a neurotoxin with major effects on the developing brain. *Pediatr Res.* 2019;85(2):183–90. <https://doi.org/10.1038/s41390-018-0224-4>.
32. Falcao AS, Silva RF, Pancadas S, Fernandes A, Brito MA, Brites D. Apoptosis and impairment of neurite network by short exposure of immature rat cortical neurons to unconjugated bilirubin increase with cell differentiation and are additionally enhanced by an inflammatory stimulus. *J Neurosci Res.* 2007;85(6):1229–39. <https://doi.org/10.1002/jnr.21227>.
33. Vaz AR, Delgado-Esteban M, Brito MA, Bolanos JP, Brites D, Almeida A. Bilirubin selectively inhibits cytochrome c oxidase activity and induces apoptosis in immature cortical neurons: assessment of the protective effects of glycoconjugated deoxycholic acid. *J Neurochem.* 2010;112(1):56–65. <https://doi.org/10.1111/j.1471-4159.2009.06429.x>.
34. Bertini G, Dani C, Pezzati M, Rubaltelli FF. Prevention of bilirubin encephalopathy. *Biol Neonate.* 2001;79(3–4):219–23. <https://doi.org/10.1159/000047095>.
35. Silva R, Mata LR, Gulbenkian S, Brito MA, Tiribelli C, Brites D. Inhibition of glutamate uptake by unconjugated bilirubin in cultured cortical rat astrocytes: role of concentration and pH. *Biochem Biophys Res Commun.* 1999;265(1):67–72. <https://doi.org/10.1006/bbrc.1999.1646>.
36. Fernandes A, Silva RF, Falcao AS, Brito MA, Brites D. Cytokine production, glutamate release and cell death in rat cultured astrocytes treated with unconjugated bilirubin and LPS. *J Neuroimmunol.* 2004;153(1–2):64–75. <https://doi.org/10.1016/j.jneuroim.2004.04.007>.
37. Grojean S, Koziel V, Vert P, Daval JL. Bilirubin induces apoptosis via activation of NMDA receptors in developing rat brain neurons. *Exp Neurol.* 2000;166(2):334–41. <https://doi.org/10.1006/exnr.2000.7518>.
38. Grojean S, Lievre V, Koziel V, Vert P, Daval JL. Bilirubin exerts additional toxic effects in hypoxic cultured neurons from the developing rat brain by the recruitment of glutamate neurotoxicity. *Pediatr Res.* 2001;49(4):507–13. <https://doi.org/10.1203/00006450-200104000-00012>.
39. McDonald JW, Shapiro SM, Silverstein FS, Johnston MV. Role of glutamate receptor-mediated excitotoxicity in bilirubin-induced brain injury in the Gunn rat model. *Exp Neurol.* 1998;150(1):21–9. <https://doi.org/10.1006/exnr.1997.6762>.
40. Mattson MP. Excitotoxic and excitoprotective mechanisms: abundant targets for the prevention and treatment of neurodegenerative disorders. *Neuromol Med.* 2003;3(2):65–94. <https://doi.org/10.1385/NMM:3:2:65>.
41. Rodrigues CM, Sola S, Castro RE, Laires PA, Brites D, Moura JJ. Perturbation of membrane dynamics in nerve cells as an early event during bilirubin-induced apoptosis. *J Lipid Res.* 2002;43(6):885–94.
42. Rodrigues CM, Sola S, Brites D. Bilirubin induces apoptosis via the mitochondrial pathway in developing rat brain neurons. *Hepatology.* 2002;35(5):1186–95. <https://doi.org/10.1053/jhep.2002.32967>.
43. Seubert JM, Darmon AJ, El-Kadi AO, D'Souza SJ, Bend JR. Apoptosis in murine hepatoma Hepa 1c1c7 wild-type, C12, and C4 cells mediated by bilirubin. *Mol Pharmacol.* 2002;62(2):257–64. <https://doi.org/10.1124/mol.62.2.257>.
44. Ahdab-Barmada M. Kernicterus in the premature neonate. *J Perinatol.* 1987;7(2):149–52.
45. Ahdab-Barmada M, Moossy J. The neuropathology of kernicterus in the premature neonate: diagnostic problems. *J Neuropathol Exp Neurol.* 1984;43(1):45–56. <https://doi.org/10.1097/00005072-198401000-00004>.
46. Conlee JW, Shapiro SM. Morphological changes in the cochlear nucleus and nucleus of the trapezoid body in Gunn rat pups. *Hear Res.* 1991;57(1):23–30. [https://doi.org/10.1016/0378-5955\(91\)90070-p](https://doi.org/10.1016/0378-5955(91)90070-p).
47. Shapiro SM, Conlee JW. Brainstem auditory evoked potentials correlate with morphological changes in Gunn rat pups. *Hear Res.* 1991;57(1):16–22. [https://doi.org/10.1016/0378-5955\(91\)90069-1](https://doi.org/10.1016/0378-5955(91)90069-1).
48. Conlee JW, Shapiro SM. Development of cerebellar hypoplasia in jaundiced Gunn rats: a quantitative light microscopic analysis. *Acta Neuropathol.* 1997;93(5):450–60. <https://doi.org/10.1007/s004010050639>.
49. Conlee JW, Shapiro SM, Churn SB. Expression of the alpha and beta subunits of Ca²⁺/calmodulin kinase II in the cerebellum of jaundiced Gunn rats during development: a quantitative light microscopic analysis. *Acta Neuropathol.* 2000;99(4):393–401. <https://doi.org/10.1007/s004010051141>.
50. Shaia WT, Shapiro SM, Heller AJ, Galiani DL, Sismanis A, Spencer RF. Immunohistochemical localization of calcium-binding proteins in the brainstem vestibular nuclei of the jaundiced Gunn rat. *Hear Res.* 2002;173(1–2):82–90. [https://doi.org/10.1016/s0378-5955\(02\)00631-7](https://doi.org/10.1016/s0378-5955(02)00631-7).
51. Spencer RF, Shaia WT, Gleason AT, Sismanis A, Shapiro SM. Changes in calcium-binding protein expression in the auditory brainstem nuclei of the jaundiced Gunn rat. *Hear Res.* 2002;171(1–2):129–41. [https://doi.org/10.1016/s0378-5955\(02\)00494-x](https://doi.org/10.1016/s0378-5955(02)00494-x).
52. Shaia WT, Shapiro SM, Spencer RF. The jaundiced Gunn rat model of auditory neuropathy/dyssynchrony. *Laryngoscope.*

- 2005;115(12):2167–73. <https://doi.org/10.1097/01.MLG.0000181501.80291.05>.
53. Zhang L, Liu W, Tanswell AK, Luo X. The effects of bilirubin on evoked potentials and long-term potentiation in rat hippocampus in vivo. *Pediatr Res.* 2003;53(6):939–44. <https://doi.org/10.1203/01.PDR.0000061563.63230.86>.
 54. Gazzin S, Zelenka J, Zdrahalova L, Konickova R, Zabetta CC, Giraudi PJ, et al. Bilirubin accumulation and Cyp mRNA expression in selected brain regions of jaundiced Gunn rat pups. *Pediatr Res.* 2012;71(6):653–60. <https://doi.org/10.1038/pr.2012.23>.
 55. Hegeman DJ, Hong ES, Hernandez VM, Chan CS. The external globus pallidus: progress and perspectives. *Eur J Neurosci.* 2016;43(10):1239–65. <https://doi.org/10.1111/ejn.13196>.
 56. Kita H. Parvalbumin-immunopositive neurons in rat globus pallidus: a light and electron microscopic study. *Brain Res.* 1994;657(1–2):31–41. [https://doi.org/10.1016/0006-8993\(94\)90950-4](https://doi.org/10.1016/0006-8993(94)90950-4).
 57. Mallet N, Micklem BR, Henny P, Brown MT, Williams C, Bolam JP, et al. Dichotomous organization of the external globus pallidus. *Neuron.* 2012;74(6):1075–86. <https://doi.org/10.1016/j.neuron.2012.04.027>.
 58. Nobrega-Pereira S, Gelman D, Bartolini G, Pla R, Pierani A, Marin O. Origin and molecular specification of globus pallidus neurons. *J Neurosci.* 2010;30(8):2824–34. <https://doi.org/10.1523/JNEUROSCI.4023-09.2010>.
 59. Borst P, Elferink RO. Mammalian ABC transporters in health and disease. *Annu Rev Biochem.* 2002;71:537–92. <https://doi.org/10.1146/annurev.biochem.71.102301.093055>.
 60. Johnson AD, Kavousi M, Smith AV, Chen MH, Dehghan A, Aspelund T, et al. Genome-wide association meta-analysis for total serum bilirubin levels. *Hum Mol Genet.* 2009;18(14):2700–10. <https://doi.org/10.1093/hmg/ddp202>.
 61. Watchko JF, Lin Z. Exploring the genetic architecture of neonatal hyperbilirubinemia. *Semin Fetal Neonatal Med.* 2010;15(3):169–75. <https://doi.org/10.1016/j.siny.2009.11.003>.
 62. Brites D. The evolving landscape of neurotoxicity by unconjugated bilirubin: role of glial cells and inflammation. *Front Pharmacol.* 2012;3:88. <https://doi.org/10.3389/fphar.2012.00088>.
 63. Fernandes A, Falcao AS, Silva RF, Brito MA, Brites D. MAPKs are key players in mediating cytokine release and cell death induced by unconjugated bilirubin in cultured rat cortical astrocytes. *Eur J Neurosci.* 2007;25(4):1058–68. <https://doi.org/10.1111/j.1460-9568.2007.05340.x>.
 64. Fernandes A, Falcao AS, Silva RF, Gordo AC, Gama MJ, Brito MA, et al. Inflammatory signalling pathways involved in astroglial activation by unconjugated bilirubin. *J Neurochem.* 2006;96(6):1667–79. <https://doi.org/10.1111/j.1471-4159.2006.03680.x>.
 65. Barateiro A, Miron VE, Santos SD, Relvas JB, Fernandes A, Ffrench-Constant C, et al. Unconjugated bilirubin restricts oligodendrocyte differentiation and axonal myelination. *Mol Neurobiol.* 2013;47(2):632–44. <https://doi.org/10.1007/s12035-012-8364-8>.
 66. Barateiro A, Vaz AR, Silva SL, Fernandes A, Brites D. ER stress, mitochondrial dysfunction and calpain/JNK activation are involved in oligodendrocyte precursor cell death by unconjugated bilirubin. *Neuromol Med.* 2012;14(4):285–302. <https://doi.org/10.1007/s12017-012-8187-9>.
 67. Riordan SM, Bittel DC, Le Pichon JB, Gazzin S, Tiribelli C, Watchko JF, et al. A hypothesis for using pathway genetic load analysis for understanding complex outcomes in bilirubin encephalopathy. *Front Neurosci.* 2016;10:376. <https://doi.org/10.3389/fnins.2016.00376>.
 68. Maisels MJ, Bhutani VK, Bogen D, Newman TB, Stark AR, Watchko JF. Hyperbilirubinemia in the newborn infant > or =35 weeks' gestation: an update with clarifications. *Pediatrics.* 2009;124(4):1193–8. <https://doi.org/10.1542/peds.2009-0329>.
 69. Cat FC, Cat A, Cicek T, Gulec SG. Evaluation of the relationship between transcutaneous bilirubin measurement and total serum bilirubin in neonatal patients followed for jaundice. *Sisli Etfal Hastan Tip Bul.* 2021;55(2):262–7. <https://doi.org/10.14744/SEMB.2020.79837>.
 70. Khan DS, Mirza A, Bhatti A, Shabbir A, Tariq B, Rizvi A. Effectiveness of transcutaneous bilirubin measurement in high-risk neonates and to evaluate validity of transcutaneous bilirubin with total serum bilirubin levels in both low and high-risk neonates at a tertiary care center in a developing country. *Cureus.* 2021;13(3):e13685. <https://doi.org/10.7759/cureus.13685>.
 71. Jegathesan T, Campbell DM, Ray JG, Shah V, Berger H, Hayeems RZ, et al. Transcutaneous versus total serum bilirubin measurements in preterm infants. *Neonatology.* 2021;118(4):443–53. <https://doi.org/10.1159/000516648>.
 72. Panda SK, Gaurav A, Das P, Swain N, Rath S. A comparison between transcutaneous bilirubin and total serum bilirubin levels for the management of jaundice in preterm neonates by Bland-Altman plot. *Cureus.* 2021;13(10):e18442. <https://doi.org/10.7759/cureus.18442>.
 73. Kumar D, Kumar D. Can serum albumin level affect the transcutaneous bilirubinometry in term neonates? *J Neonatal Perinatal Med.* 2022. <https://doi.org/10.3233/NPM-210958>.
 74. Ho SR, Lin YC, Chen CN. The impact of phototherapy on the accuracy of transcutaneous bilirubin measurements in neonates: optimal measurement site and timing. *Diagnostics (Basel).* 2021;11(9). <https://doi.org/10.3390/diagnostics11091729>.
 75. Olusanya BO, Slusher TM, Imosemi DO, Emokpae AA. Maternal detection of neonatal jaundice during birth hospitalization using a novel two-color icterometer. *PLoS ONE.* 2017;12(8):e0183882. <https://doi.org/10.1371/journal.pone.0183882>.
 76. Taylor JA, Stout JW, de Greef L, Goel M, Patel S, Chung EK, et al. Use of a smartphone app to assess neonatal jaundice. *Pediatrics.* 2017;140(3). <https://doi.org/10.1542/peds.2017-0312>.
 77. Hegyi T, Kleinfeld A, Huber A, Weinberger B, Memon N, Shih W, et al. Unbound bilirubin measurements by a novel probe in preterm infants. *J Matern Fetal Neonatal Med.* 2019;32(16):2721–6. <https://doi.org/10.1080/14767058.2018.1448380>.
 78. Huber AH, Zhu B, Kwan T, Kampf JP, Hegyi T, Kleinfeld AM. Fluorescence sensor for the quantification of unbound bilirubin concentrations. *Clin Chem.* 2012;58(5):869–76. <https://doi.org/10.1373/clinchem.2011.176412>.
 79. Johnson L, Brown AK, Bhutani VK. BIND - a clinical score for bilirubin induced neurologic dysfunction in newborns. *Pediatrics.* 1999;104(3):746–7.
 80. Radmacher PG, Groves FD, Owa JA, Ofowwe GE, Amuabunos EA, Olusanya BO, et al. A modified bilirubin-induced neurologic dysfunction (BIND-M) algorithm is useful in evaluating severity of jaundice in a resource-limited setting. *BMC Pediatr.* 2015;15:28. <https://doi.org/10.1186/s12887-015-0355-2>.
 81. Dasari VR, Shapiro SM, Yeh HW, Gelineau-Morel R. Kernicterus Spectrum Disorders Diagnostic Toolkit: validation using retrospective chart review. *Pediatr Res.* 2021. <https://doi.org/10.1038/s41390-021-01755-5>.
 82. Coskun A, Yikilmaz A, Kumandas S, Karahan OI, Akcakus M, Manav A. Hyperintense globus pallidus on T1-weighted MR imaging in acute kernicterus: is it common or rare? *Eur Radiol.* 2005;15(6):1263–7. <https://doi.org/10.1007/s00330-004-2502-2>.

83. Wang X, Wu W, Hou BL, Zhang P, Chineah A, Liu F, et al. Studying neonatal bilirubin encephalopathy with conventional MRI, MRS, and DWI. *Neuroradiology*. 2008;50(10):885–93. <https://doi.org/10.1007/s00234-008-0423-5>.
84. Mao J, Fu JH, Chen LY, Wang XM, Xue XD. Changes of globus pallidus in the newborn infants with severe hyperbilirubinemia. *Zhonghua Er Ke Za Zhi*. 2007;45(1):24–9.
85. Cece H, Abuhandan M, Cakmak A, Yildiz S, Calik M, Karakas E, et al. Diffusion-weighted imaging of patients with neonatal bilirubin encephalopathy. *Jpn J Radiol*. 2013;31(3):179–85. <https://doi.org/10.1007/s11604-012-0166-4>.
86. Wu M, Shen X, Lai C, You Y, Zhao Z, Wu D. Detecting acute bilirubin encephalopathy in neonates based on multimodal MRI with deep learning. *Pediatr Res*. 2021. <https://doi.org/10.1038/s41390-021-01560-0>.
87. Wu W, Zhang P, Wang X, Chineah A, Lou M. Usefulness of (1) H-MRS in differentiating bilirubin encephalopathy from severe hyperbilirubinemia in neonates. *J Magn Reson Imaging*. 2013;38(3):634–40. <https://doi.org/10.1002/jmri.23995>.
88. Wisnowski JL, Panigrahy A, Painter MJ, Watchko JF. Magnetic resonance imaging of bilirubin encephalopathy: current limitations and future promise. *Semin Perinatol*. 2014;38(7):422–8. <https://doi.org/10.1053/j.semperi.2014.08.005>.
89. Stevenson DK, Wong RJ, Arnold CC, Pedroza C, Tyson JE. Phototherapy and the risk of photo-oxidative injury in extremely low birth weight infants. *Clin Perinatol*. 2016;43(2):291–5. <https://doi.org/10.1016/j.clp.2016.01.005>.
90. Lightner DA, Linnane WP 3rd, Ahlfors CE. Bilirubin photooxidation products in the urine of jaundiced neonates receiving phototherapy. *Pediatr Res*. 1984;18(8):696–700. <https://doi.org/10.1203/00006450-198408000-00003>.
91. Hansen TW. The role of phototherapy in the crash-cart approach to extreme neonatal jaundice. *Semin Perinatol*. 2011;35(3):171–4. <https://doi.org/10.1053/j.semperi.2011.02.012>.
92. Olusanya BO, Imam ZO, Emokpae AA, Iskander IF. Revisiting the criteria for exchange transfusion for severe neonatal hyperbilirubinemia in resource-limited settings. *Neonatology*. 2016;109(2):97–104. <https://doi.org/10.1159/000441324>.
93. Maisels MJ, Watchko JF, Bhutani VK, Stevenson DK. An approach to the management of hyperbilirubinemia in the preterm infant less than 35 weeks of gestation. *J Perinatol*. 2012;32(9):660–4. <https://doi.org/10.1038/jp.2012.71>.
94. Murki S, Kumar P. Blood exchange transfusion for infants with severe neonatal hyperbilirubinemia. *Semin Perinatol*. 2011;35(3):175–84. <https://doi.org/10.1053/j.semperi.2011.02.013>.
95. Gottstein R, Cooke RW. Systematic review of intravenous immunoglobulin in haemolytic disease of the newborn. *Arch Dis Child Fetal Neonatal Ed*. 2003;88(1):F6–10. <https://doi.org/10.1136/fn.88.1.f6>.
96. Suresh GK, Martin CL, Soll RF. Metalloporphyrins for treatment of unconjugated hyperbilirubinemia in neonates. *Cochrane Database Syst Rev*. 2003;(2):CD004207. <https://doi.org/10.1002/14651858.CD004207>.
97. Martinez JC, Garcia HO, Otheguy LE, Drummond GS, Kappas A. Control of severe hyperbilirubinemia in full-term newborns with the inhibitor of bilirubin production Sn-mesoporphyrin. *Pediatrics*. 1999;103(1):1–5. <https://doi.org/10.1542/peds.103.1.1>.
98. Kappas A, Drummond GS, Henschke C, Valaes T. Direct comparison of Sn-mesoporphyrin, an inhibitor of bilirubin production, and phototherapy in controlling hyperbilirubinemia in term and near-term newborns. *Pediatrics*. 1995;95(4):468–74.
99. Kappas A, Drummond GS, Munson DP, Marshall JR. Sn-Mesoporphyrin interdiction of severe hyperbilirubinemia in Jehovah's Witness newborns as an alternative to exchange transfusion. *Pediatrics*. 2001;108(6):1374–7. <https://doi.org/10.1542/peds.108.6.1374>.
100. Rice AC, Chiou VL, Zuckoff SB, Shapiro SM. Profile of minocycline neuroprotection in bilirubin-induced auditory system dysfunction. *Brain Res*. 2011;1368:290–8. <https://doi.org/10.1016/j.brainres.2010.10.052>.
101. Vodret S, Bortolussi G, Iaconcig A, Martinelli E, Tiribelli C, Muro AF. Attenuation of neuro-inflammation improves survival and neurodegeneration in a mouse model of severe neonatal hyperbilirubinemia. *Brain Behav Immun*. 2018;70:166–78. <https://doi.org/10.1016/j.bbi.2018.02.011>.
102. Bortolussi G, Baj G, Vodret S, Viviani G, Bittolo T, Muro AF. Age-dependent pattern of cerebellar susceptibility to bilirubin neurotoxicity in vivo in mice. *Dis Model Mech*. 2014;7(9):1057–68. <https://doi.org/10.1242/dmm.016535>.
103. Deliktas M, Ergin H, Demiray A, Akca H, Ozdemir OMA, Ozdemir MB. Caffeine prevents bilirubin-induced cytotoxicity in cultured newborn rat astrocytes. *J Matern Fetal Neonatal Med*. 2019;32(11):1813–9. <https://doi.org/10.1080/14767058.2017.1419175>.
104. Kuter N, Aysit-Altuncu N, Ozturk G, Ozek E. The neuroprotective effects of hypothermia on bilirubin-induced neurotoxicity in vitro. *Neonatology*. 2018;113(4):360–5. <https://doi.org/10.1159/000487221>.
105. De Siati RD, Rosenzweig F, Gersdorff G, Gregoire A, Rombaux P, Deggouj N. Auditory neuropathy spectrum disorders: from diagnosis to treatment: literature review and case reports. *J Clin Med*. 2020;9(4). <https://doi.org/10.3390/jcm9041074>.
106. Shapiro SM, Riordan SM. Review of bilirubin neurotoxicity II: preventing and treating acute bilirubin encephalopathy and kernicterus spectrum disorders. *Pediatr Res*. 2020;87(2):332–7. <https://doi.org/10.1038/s41390-019-0603-5>. **This review summarizes current and possible novel methods to prevent bilirubin neurotoxicity and treat ABE and KSDs.**
107. Sanger TD, Liker M, Arguelles E, Deshpande R, Maskooki A, Ferman D, et al. Pediatric deep brain stimulation using awake recording and stimulation for target selection in an inpatient neuromodulation monitoring unit. *Brain Sci*. 2018;8(7). <https://doi.org/10.3390/brainsci8070135>.
108. Yuan H, Li Y, Yang J, Li H, Yang Q, Guo C, et al. State of the art of non-invasive electrode materials for brain-computer interface. *Micromachines (Basel)*. 2021;12(12). <https://doi.org/10.3390/mi12121521>.
109. Foldes ST, Chandrasekaran S, Camerone J, Lowe J, Ramdeo R, Ebersole J, et al. Case study: mapping evoked fields in primary motor and sensory areas via magnetoencephalography in tetraplegia. *Front Neurol*. 2021;12:739693. <https://doi.org/10.3389/fneur.2021.739693>.
110. Oxley TJ, Yoo PE, Rind GS, Ronayne SM, Lee CMS, Bird C, et al. Motor neuroprosthesis implanted with neurointerventional surgery improves capacity for activities of daily living tasks in severe paralysis: first in-human experience. *J Neurointerv Surg*. 2021;13(2):102–8. <https://doi.org/10.1136/neurintsurg-2020-016862>.
111. Yang FC, Riordan SM, Winter M, Gan L, Smith PG, Vivian JL, et al. Fate of neural progenitor cells transplanted into jaundiced and nonjaundiced rat brains. *Cell Transplant*. 2017;26(4):605–11. <https://doi.org/10.3727/096368917X694840>.
112. Yang FC, Draper J, Smith PG, Vivian JL, Shapiro SM, Stanford JA. Short term development and fate of MGE-like neural

- progenitor cells in jaundiced and non-jaundiced rat brain. *Cell Transplant*. 2018;27(4):654–65. <https://doi.org/10.1177/0963689718766327>.
113. Amini N, Vousooghi N, Soleimani M, Samadikuchaksaraei A, Akbari M, Safakheil H, et al. A new rat model of neonatal bilirubin encephalopathy (kernicterus). *J Pharmacol Toxicol Methods*. 2017;84:44–50. <https://doi.org/10.1016/j.vascn.2016.10.002>.
114. Amini N, Vousooghi N, Hadjighassem M, Bakhtiyari M, Mousavi N, Safakheil H, et al. Efficacy of human adipose tissue-derived stem cells on neonatal bilirubin encephalopathy in rats. *Neurotox Res*. 2016;29(4):514–24. <https://doi.org/10.1007/s12640-016-9599-3>.

Publisher's Note Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.